

Protein-induced morphological transitions in KCl crystal growth

Bálint Szabó and Tamás Vicsek

Department of Biological Physics, Eötvös University, Budapest, Pázmány Péter sétány 1/A, 1117 Hungary

(Received 16 August 2002; published 24 January 2003)

We investigated the formation of KCl crystals on a glass surface by phase contrast, fluorescent, and atomic force microscopy on the micrometer scale and observed interesting morphological transitions as a function of the experimental conditions. The presence of proteins in the solution from which the salt crystals grow during the drying up leads to complex microscopic patterns of crystals, some of which are analogous to those commonly observed on the macroscopic scale. We tested the effect of tubulin, FITC-labeled albumin, and IgG on the morphology of crystals grown either slowly or fast. A rich variety of protein-specific and concentration-dependent morphologies was found and described by a morphological diagram. We give a phenomenological interpretation, which can explain the growth of complex patterns. Fluorescent images prove that a protein layer covers the surface of the KCl structures. We propose that this layer reduces the anisotropy of the effective surface tension during growth. The tip splitting fractal regime is attributed to the decrease of anisotropy. Further possible mechanisms, which can cause a morphological transition, are also discussed. We found elongated saw-toothed crystals induced by proteins, especially IgG, and identified their structure.

DOI: 10.1103/PhysRevE.67.011908

PACS number(s): 87.90.+y, 89.75.Kd, 47.20.Dr, 68.37.-d

I. INTRODUCTION

The effect of proteins, especially albumin and IgG, on crystal growth has biological relevance. In crystal-induced arthritis—e.g., gout—proteins bind the crystals [1] and have significant impact on crystal growth [2]. The immune response to the appearance of crystals is driven by the specific IgG-crystal interaction. This interaction stimulates crystal formation [3–5].

Although the rich morphology of several inorganic macroscopic crystals grown on the surface of gels (i.e., in the presence of protein) has been studied and discussed [6,7] to our knowledge this is the first study of pattern formation induced by crystal-protein interactions.

Moving unstable interfaces typically lead to the formation of complex patterns. Surface tension of the boundary between the growing and surrounding phases is essential in pattern formation [8–11]. A growth mode of crystals covered with a thin layer has been observed and explained by the surface tension between the different phases in a metal system [12]. The growth of nonequilibrium interfaces covered by a thin surfactant film was both experimentally and theoretically studied [13]. The anisotropy of the surface tension has a crucial role. Viscous fingering, crystallization, electrochemical deposition, and some other phenomena can be relatively well described by Laplacian growth with appropriate boundary conditions. We argue that the growth of the numerous different microscopic patterns we observed in our KCl crystallization experiments in the presence of proteins can be explained by the change of the boundary conditions of the moving interface: the anisotropy of its surface tension is decreased by proteins.

II. MATERIALS AND METHODS

5 μ l drops of 5 mg/ml KCl solution containing either tubulin (prepared from bovine brain [14]) or FITC-BSA (bovine serum albumin conjugated with FITC, SIGMA) or hu-

man IgG-FITC (immunoglobulin G conjugated with FITC, SIGMA) or no protein were placed on clean glass cover slips. We prepared samples either by slow drying at $\sim 40^\circ\text{C}$ in air or by fast drying in a strong airflow. Slow drying took ~ 5 min; airflow dried the surface on the scale of seconds. Bright field, phase contrast, and fluorescent images were acquired by an inverse Leica DM IRB microscope, $40\times$ objective, and a Nikon Coolpix 700 digital camera mounted on the microscope by homemade optical coupling. Atomic force microscopy (AFM) images of tubulin-coated crystals were captured with a commercial AFM (TopoMetrix Explorer, Santa Clara, CA) in contact mode with a soft silicon nitride cantilever (Thermomicroscopes, coated sharp microlevers, model No. MSCT-AUHW, with typical force constant 0.03 N/m, 20 nm nominal radius of curvature) under ambient conditions.

III. RESULTS**A. Tubulin**

The fast drying of 0.5 mg/ml tubulin containing drops of 5 mg/ml KCl solution on glass surface with airflow results in a heterogeneous population of patterns that we investigated by AFM and phase contrast microscopy. The morphology of KCl crystals can be similar to diffusion-limited aggregates known to have fractal geometry. Dendritic patterns with stable and unstable tips are typical. Figure 1 shows a pattern, which began to grow with stable tips, four-fold symmetry. After a while tip splitting occurs at all four tips. Anisotropy drops dramatically at this point. The slow drying of drops containing 0.05 mg/ml tubulin gives elongated saw-toothed crystals similar to the ones induced by IgG discussed below. In high concentration tubulin inhibits crystal growth; an apparently homogeneous stain can be observed after the drop dries up.

B. FITC-BSA

We tried to shed light on the role of protein in the pattern formation of salt crystals with the use of FITC-labeled pro-

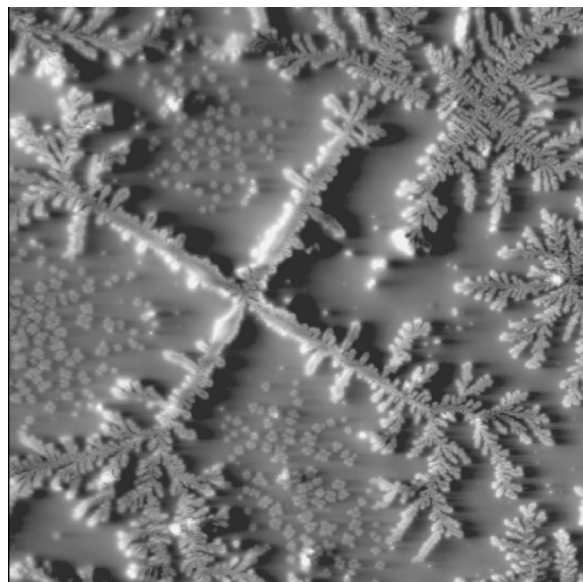


FIG. 1. $80 \times 80 \mu\text{m}^2$ deflection mode AFM image of tubulin induced patterns of KCl after the fast drying procedure. Note the transition of the initially four-fold symmetric crystallization to isotropic fractal growth at all four tips of the structure caused by consecutive tip splittings.

teins. In case of the fast drying procedure 5 mg/ml FITC-BSA had a similar effect to that of tubulin. Various branching morphologies were found. Tip splitting caused fractal growth (Fig. 2). The comparison of bright field and phase contrast images with the fluorescent ones shows that crystals are covered by a layer of albumin. The protein concentration dependence of the morphology was studied with the slow drying technique. Under a critical protein concentration



FIG. 2. $135 \times 135 \mu\text{m}^2$ fluorescent image of two patterns of KCl crystals with different fractal dimensions grown in the presence of FITC-BSA with the fast drying method. Brightness is proportional to the concentration of FITC-BSA. The comparison with phase contrast and bright field images proves that albumin covers the crystals.

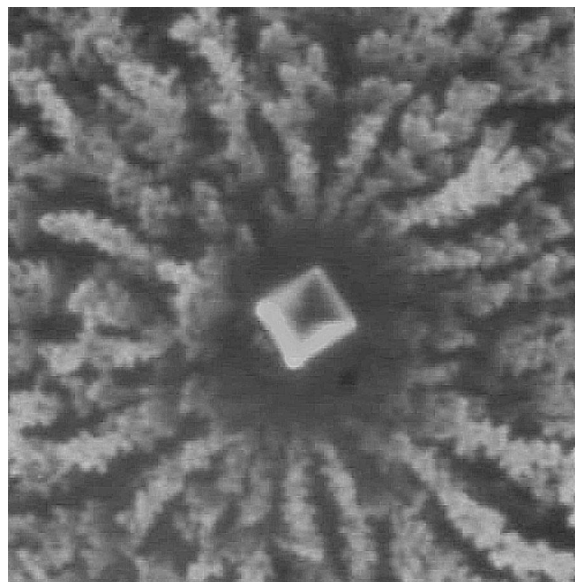


FIG. 3. $135 \times 135 \mu\text{m}^2$ fluorescent image of a cubic crystal centered structure of KCl crystals induced by FITC-BSA using the slow drying method. The anisotropic single crystal in the center is surrounded by an isotropic pattern.

($\sim 1 \mu\text{g/ml}$) only blocks of rectangular prism-shaped crystals grew corresponding to the protein-free case. In the concentration range of 1–1000 $\mu\text{g/ml}$ cubic-crystal-centered structures formed (Fig. 3). This observation reinforced that during a single growth process the symmetry (anisotropy) of the pattern can dramatically change due to the change of the local conditions. As the growth process elongates, the initial four-fold symmetry with high anisotropy disappears and rather isotropic growth takes place. In the higher-protein-concentration range dendritic crystals grew with stable tips (Fig. 4).



FIG. 4. $135 \times 135 \mu\text{m}^2$ fluorescent image of stable tipped dendritic KCl crystals induced by a high concentration of albumin.

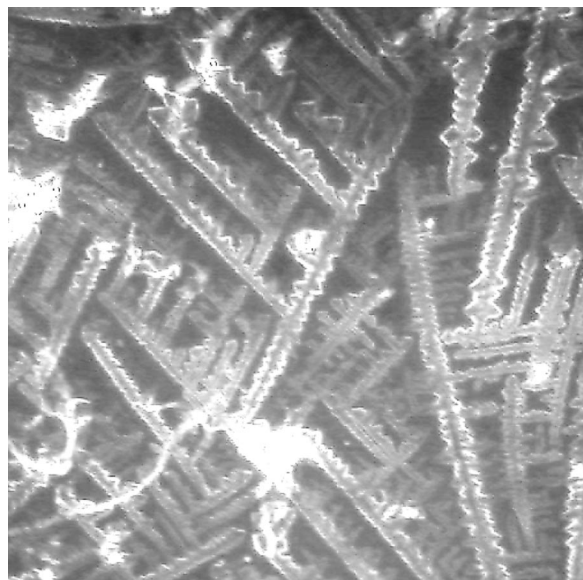


FIG. 5. $135 \times 135 \mu\text{m}^2$ fluorescent image of KCl crystals with saw-toothed shape grown with IgG by the fast drying procedure. The angle between the branches is $\sim 70^\circ$. The bright contour of the crystals corresponds to the IgG-FITC cover on their surface.

C. IgG-FITC

To study the protein specificity of the phenomena we examined the impact of human IgG-FITC on the KCl crystal growth. After the fast drying procedure using 2 mg/ml IgG the typical pattern we found was the elongated saw-toothed crystal (Fig. 5). Branching structures can be observed. The angle between the branches is $\sim 70^\circ$. The concentration dependence of the morphology was investigated by the slow drying method. Under $\sim 20 \mu\text{g/ml}$ only blocks of the prism-shaped crystals grew. Above this concentration up to $\sim 1 \text{ mg/ml}$ cubic and saw-toothed crystal-centered structures were observed with an isotropic surrounding pattern. Figure 6 shows the center of a typical one. In the higher-concentration range dendritic patterns formed with stable tips.

D. Equilibrium shape

The equilibrium shape of KCl crystals after the slow drying method using $10 \mu\text{l}$ 5 mg/ml KCl with 5 mg/ml FITC-BSA was observed by dropping $5 \mu\text{l}$ saturated KCl solution on the crystals and covering the droplet with a cover slip. We found rounded forms instead of the normal rectangular shapes of KCl.

E. Gold substrate

We also studied the effect of the substrate surface, which was glass in the above experiments. However, IgG gave similar results on gold substrate to those on glass in the case of the slow drying method; albumin induced the appearance of amorphous protein aggregates and blocks of prism-shaped crystals on gold instead of the patterns described above. This fact indicates the protein and substrate specificity of the phenomenon.

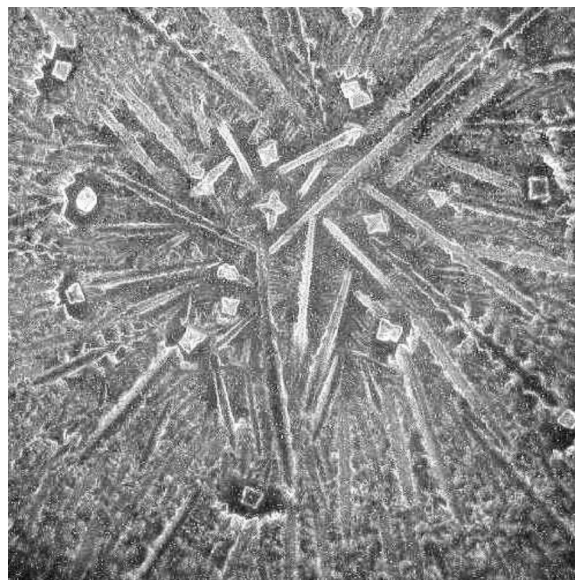


FIG. 6. $205 \times 205 \mu\text{m}^2$ fluorescent image. Saw-toothed crystals are shown in the center of a pattern, which is isotropic in the surrounding regions. This sample was prepared with the slow drying procedure in the presence of IgG.

IV. DISCUSSION

Proteins have significant impact on KCl crystallization. The presence of proteins in the solution from which KCl crystals grow during the drying up leads to the formation of protein-specific and concentration-dependent complex patterns, which we described by a morphological diagram (Fig. 7). We can give a partial explanation of the diagram.

During the drying up of the solution protein precipitates due to the increasing ionic strength. This process is known as salting out. A gradually thickening layer of protein aggregate (precipitate) covers the surface of crystals. This cover means two interfaces: one between the KCl crystal and the protein layer and one between the protein layer and the liquid. While the surface tension of the former one, γ_{c-p} , is anisotropic, that of the latter one, γ_{p-l} , is considered to be isotropic. The total surface tension γ_t of the interface between the growing and surrounding phases is given by the sum of these two terms:

$$\gamma_t = \gamma_{c-p} + \gamma_{p-l}. \quad (1)$$

The value of γ_{p-l} , i.e., the isotropic term, is significant since precipitation means insolubility of the protein. If $\gamma_{c-p} < \gamma_{p-l}$, then γ_t is dominated by the isotropic term. The rounded equilibrium shape of crystals with protein cover reinforces the lower anisotropy of surface tension. In the absence of protein the extra isotropic term corresponding to the protein-liquid interface is missing. We propose that tip splitting and fractal growth instead of the formation of single crystals with four-fold symmetry and stable tips can be attributed to the reduced anisotropy of the surface tension in the case of the fast growth. Similarly, slowly grown single-crystal-centered structures are explained by the decrease of anisotropy of surface tension during the growth process. Our

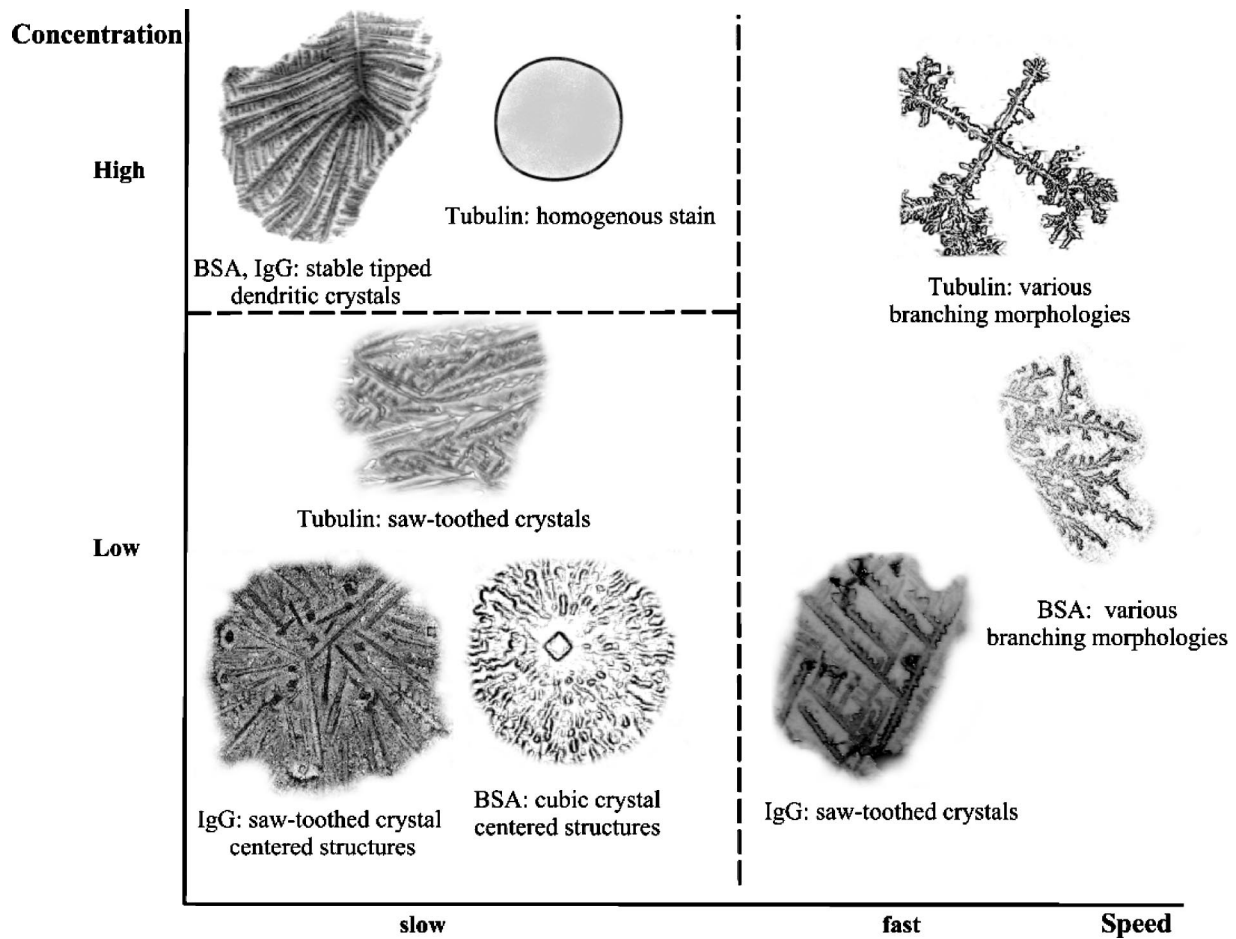


FIG. 7. Morphological diagram of the protein-induced patterns of KCl crystals.

results demonstrate that the level of anisotropy can change dramatically during the growth of a single pattern. This is likely to be caused by the increase and the reaching of a critical level of protein precipitation as the drop is drying up.

Various examples of pattern formation driven by moving unstable interfaces can be described by Laplacian growth. In this case the boundary condition along the Γ interface containing the dimensionless u_Γ concentration term is [8–11]

$$u_\Gamma = \Delta - d_0(\Theta)\kappa - \beta(\Theta)v_n, \quad (2)$$

where Δ is the undercooling, the capillary length d_0 is proportional to the surface tension, κ denotes the local curvature of the interface, β is the kinetic coefficient, Θ is the angle between the normal to the surface and a fixed crystallographic direction, and v_n is the normal velocity of the interface. To explain the stable-tip-to-tip-splitting transition, we propose that d_0 is altered and its anisotropy is decreased due to the adherence of the protein layer to the KCl crystal surface.

Further effects may also have an impact on the morphology of KCl patterns. The lower diffusion coefficient of K^+ and Cl^- ions in the liquid phase due to the high protein concentration during the drying up might induce transport-limited growth, typically resulting in fractal geometry [8]. The modification of the angular dependence of β by a more

isotropic incorporation process of KCl into the crystal caused by the protein coating can lead to a lower anisotropy of the pattern. Faster growth, which probably appears in the final state of growth, strengthens the impact of the kinetic term. The incorporation of proteins into calcite crystals is known to have an effect on the morphology [15]. Although fluorescent images displayed high protein concentration on the surface of crystals, we cannot totally exclude incorporation.

IgG typically and under some conditions tubulin and albumin also induced the growth of elongated saw-toothed crystals instead of rectangular needle-shaped ones. Branching saw-toothed structures were also found; the angle between the branches is $\sim 70^\circ$. We argue that these are single crystals elongated in the (111) direction of the cubic crystal; i.e., their axis lies in this direction. The angle between (111) and directions with the same symmetry, e.g., (11-1), is 70.5° . The mechanism leading to the formation of these crystals is unknown. Proteins can adhere to specific faces of the crystal with increased affinity [2]. This anisotropic interaction influences the anisotropy of the surface tension at the crystal-protein interface γ_{c-p} , which may result in the change of the direction of the fast growing tip.

The background of the protein specific behavior is unclear. As the experiment on the gold substrate indicates, in addition to the protein-crystal interaction, protein-substrate and crystal-substrate effects also should be considered in further studies.

ACKNOWLEDGMENTS

This work was supported by the Hungarian National Scientific Research Fund (OTKA, Grant No. T-034995). We

thank Noémi Rozlosnik and András Czirók for the help in establishing the instrumental background of our experiments and Judit Ovádi and Emma Hlavanda for the tubulin samples.

-
- [1] O. Antommattei, R. Heimer, D.G. Baker, and H.R. Schumacher, Jr., *Clin. Exp. Rheumatol.* **8**, 29 (1990).
- [2] D. Perl-Treves and L. Addadi, *Proc. R. Soc. London, Ser. B* **235**, 145 (1988).
- [3] E. Ortiz-Bravo, M.S. Sieck, and H.R. Schumacher, Jr., *Arthritis Rheum.* **36**(9), 1274 (1993).
- [4] M. Kam, D. Perl-Treves, D. Caspi, and L. Addadi, *FASEB J.* **6**, 2608 (1992).
- [5] M. Kam, D. Perl-Treves, R. Sfez, and L. Addadi, *J. Mol. Recognit.* **7**, 257 (1994).
- [6] M. Yasui and M. Matsushita, *J. Phys. Soc. Jpn.* **61**, 2327 (1992).
- [7] J. Suda and M. Matsushita, *J. Phys. Soc. Jpn.* **64**, 348 (1995).
- [8] T. Vicsek, *Fractal Growth Phenomena*, 2nd ed. (World Scientific, London, 1992), pp. 271–339.
- [9] D.A. Kessler, J. Koplik, and H. Levine, *Adv. Phys.* **37**, 255 (1988).
- [10] E. Ben-Jacob and P. Garik, *Nature (London)* **343**, 523 (1990).
- [11] E. Ben-Jacob, N. Goldenfeld, J.S. Langer, and G. Schön, *Phys. Rev. Lett.* **51**, 1930 (1983).
- [12] C. Nagl, E. Platzgummer, M. Schmid, P. Varga, S. Speller, and W. Heiland, *Phys. Rev. Lett.* **75**, 2976 (1995).
- [13] A-L. Barabási, *Phys. Rev. Lett.* **70**, 4102 (1993).
- [14] G.C. Na and S.N. Timasheff, *Biochemistry* **25**, 6214 (1986).
- [15] S. Albeck, I. Addadi, and S. Weiner, *Connect. Tissue Res.* **35**, 365 (1996).